INTRODUCTION

Sleep occupies about a third of our lives and is essential for health and well-being [1]. Sleep disturbance adversely affects physiological, psychological, and cognitive functions. Obstructive sleep apnea (OSA) is a common sleep related breathing disorder characterized by intermittent upper-airway collapse during sleep that affects about 10%–20% of middle- and older-aged adults [2-4]. OSA makes it difficult to fall or stay asleep, resulting in poor sleep quality and sleep fragmentation [5], possibly promoting cognitive decline and neurodegeneration [6-8]. In zebrafish, hypoxia, a hallmark of OSA, induced cognitive impairment [9,10]. However, the mechanism by which sleep disorders cause cognitive dysfunction is unclear.

The hexosamine biosynthesis pathway (HBP) is active in most eukaryotic cells, accounts for 2–5% of glucose metabolism in humans, and causes intracellular protein modifications [11,12]. O-linked N-acetylglucosamine (O-GlcNAc) is attached to intracellular proteins via UDP-N-acetylglucosamine (UDP-GlcNAc), the final metabolite of HBP, and this modification (i.e., O-GlcNAcylation) promotes various cellular processes, including cell signaling, gene expression, and protein stability [12-14]. However, the role of O-GlcNAc in sleep disorders and cognitive dysfunction remains unclear.

The objective of this preliminary study was to investigate the relationships among objective sleep variables, cognitive function, and HBP activation levels in middle- and older-aged adults with/without obstructive sleep apnea (OSA).

Methods: A twenty-four participants (16 males and 8 females; mean age, 58.63±7.61 years) were recruited. The comprehensive attention test (CAT), peripheral blood collection, actigraphy (AG) and nocturnal polysomnography (nPSG) tests were conducted to evaluate cognitive function and the activation of HBP and sleep variables, respectively. Results: Total sleep time (TST) on AG was correlated positively with number of errors and mean response time (RT) on CAT. Sleep efficiency (SE) on AG was correlated negatively with number of errors and positively with mean RT, whereas number of awakenings (NOA) was inversely associated with these variables. TST and SE on nPSG, SE on AG were positively correlated with levels of hexosamine pathway metabolites, whereas wake after sleep onset and sleep latency (SL) on nPSG, SL and NOA on AG were negatively correlated therewith. There was a negative correlation between levels of hexosamine pathway metabolites and number of errors on CAT. Conclusion: Sleep variables, cognitive function (i.e., attention), and HBP activation level were significantly correlated in middle- and older-aged OSA participants.
ation) increases protein stability and prevents protein aggregation [13]. Individuals with sleep disturbance and its risk factors (e.g., sleep deprivation, OSA, shift work) tend to show increased reactive oxygen species (ROS) levels [14–16]. ROS increase insulin resistance and decrease intracellular glucose levels, leading to reduction of HBP activation and O-GlcNAcylation [17]. There is also a close relationship between sleep disorders, including OSA, and increased insulin resistance [18,19].

Decreased HBP activation may play an important role in cognitive impairment with aging. For example, O-GlcNAcylation has effects on many diseases related to aging, including diabetes, cardiomyopathies, and cancer [20]. It has been hypothesized that reduced O-GlcNAcylation in the brain causes aggregation of tau protein and accumulation of β-amyloid, which are associated with Alzheimer’s disease [21]. In addition, patients with mild cognitive impairment showed low glucose metabolism in the brain [22], and glucose in the brain blood flow decreased with aging in normal controls [23]. Furthermore, HBP activation is likely involved in the associations of various sleep disorders with cognitive dysfunction. A zebrafish study found not only learning and memory impairments but also suppressed O-GlcNAcylation after sleep deprivation [24]. To summarize, sleep disturbance may affect insulin resistance and glucose metabolism, which could inhibit HBP and ultimately contribute to reduced brain function. Thus, it is possible that HBP activation levels mediate the relationship between sleep variables and cognitive function.

We compared the HBP activation of OSA and normal control (NC) groups and investigated the relationships among sleep parameters, HBP activation level, and cognitive function in OSA patients. To the best of our knowledge, few studies have investigated the relationships among these factors in people with/without OSA. Therefore, we recruited middle- and older-aged adults and analyzed sleep parameters using objective devices [i.e., nocturnal polysomnography (nPSG) and actigraphy (AG)], HBP activation data, and cognitive function as assessed by the comprehensive attention test (CAT). We aimed to identify the mechanism by which sleep disturbance causes cognitive decline and to provide evidence for the importance of treating sleep disorders.

METHODS

Participants
Sixteen males (mean age 59.69±8.56 years, range 42–74 years) and eight females (mean age 56.50±5.04 years, range 51–65 years) were recruited from advertisements or following an outpatient visit to Seoul National University Hospital. All participants were able to understand the explanation of the experiment and had a clinical dementia rating of 0. The exclusion criteria were: major mental illnesses other than dementia, such as schizophrenia, bipolar disorder, major depression, alcohol/substance abuse or dependence, or delirium; neurological or medical disease that affects mental state; illiteracy sufficient to interfere with test performance; impairment of vision or hearing sufficient to interfere with test performance; participant in a drug trial; and other sleep disorders. This study was approved by the Institutional Review Board of Seoul National University Hospital (approval no. 1806-167-954), and all procedures were conducted after obtaining written informed consent from participants.

Materials
Participants’ demographic information including age, sex, usual sleep habits, medical history, and drug use were recorded, followed by completion of the CAT and peripheral blood collection. Participants wore an AG on their non-dominant wrist for approximately 1 week and subsequently underwent one-night of nPSG.

CAT
We conducted the CAT, a computer-based test standardized in Korea, to evaluate cognitive function [25]. Participants conducted two types of tests measuring different attention abilities: the inhibition-sustained attention test (ISAT) and the divided-attention test (DAT). Inhibition-sustained attention is the maintenance or suppression of a response to repeated stimuli or actions. To evaluate whether the inhibition of a specific stimulus was maintained, participants were instructed to respond by pressing the space bar whenever various stimuli (e.g., circles, triangles, squares) appeared on a display, except for one stimulus (i.e., X-shape). Divided attention is the processing of more than one stimulus or piece of information simultaneously. To evaluate their ability to deal with successive stimuli presented simultaneously, participants were instructed to respond by pressing the space bar whenever they judged that the same stimulus as in the previous trial had been presented, irrespective of the stimulus type (e.g., visual stimuli: circles, triangles, and squares; auditory stimuli: bell sound and beep).

All participants performed the test after sufficient practice to learn the rules. The test time was 10 minutes for ISAT and 3 minutes and 20 seconds for DAT. The following indicators were measured in each test: commission error (CE; the number of wrong responses to a non-target stimulus), omission error (OE; the number of misses without responding to the target stimulus), mean response time of correct responses (mean RT), and sensitivity coefficient (d’; an indicator of how well the target stimulus and the non-target stimulus are distinguished).

Quantitative analysis of metabolites by liquid chromatography-mass spectrometry
To evaluate HBP activation, approximately 10 mL of blood were collected from the peripheral veins of the participants by trained experts. For targeted profiling of hexosamine pathway metabolites, 50 µL of serum was mixed with 500 µL of chloroform/methanol (2:1, v/v) and 100 µL of water. The mixture was vortexed and incubated at 4°C for 10 minutes. After centrifugation, 200 µL of the supernatant was dried using N2 gas. The extracts were diluted with 50 µL of an acetonitrile/water mixture (20:80, v/v). To quantify the levels of metabolites, liquid chromatography-mass spec-
rhythm (LC/MS) was performed on an Agilent 1290 Infinity LC and an Agilent 6495 Triple Quadrupole MS system equipped with an Agilent Jet Stream ESI source (Agilent Technologies, Santa Clara, CA, USA). MassHunter Workstation (ver. B.06.00, Agilent Technologies) software was used for data acquisition and analysis. LC separation was carried out on an Intrada amino acid column (150 mm×3 mm, Imtakt, Portland, OR, USA). The binary gradient system comprised 0.3% formic acid in ACN (solvent A) and 100 mM ammonium formate in a mixture of ACN and water (20:80, v/v, solvent B). The linear gradients used for elution and equilibrating the initial gradient for subsequent runs were as follows: 0–27% B at 0–3 minutes, 27–100% B at 3–19.5 minutes, 100% B at 19.5–25 minutes, and 0% B at 25–32 minutes. The column temperature and flow rate were 25°C and 0.6 mL/minute, respectively. The autosampler temperature was maintained at 10°C, and the sample injection volume was 5 μL. Samples were analyzed in single reaction monitoring (SRM) positive ion mode. The SRM transitions were performed with the following operational parameters: capillary voltage, 2 kV; cone voltage, 20 V; source temperature, 150°C; desolvation temperature, 650°C; cone gas flow, 150 L/hour; desolvation gas flow, 900 L/hour; collision gas flow, 0.15 mL/minute; and nebulizer gas flow, 7 bar. Concentrations of metabolites were quantified based on a series of external calibration curves generated using authentic standards. As a result of LC-MS analysis, fructose-6-phosphate (F6P) and glucosamine-6-phosphate (G6P), N-acetyl glucosamine-1-phosphate (GlcNAc1P) and N-acetyl glucosamine-6-phosphate (GlcNAc6P), and UDP-GlcNAc were quantified. Two substrates were reported as one value if isomeric metabolites were not well separated (i.e., F6P and G6P, GlcNAc1P and GlcNAc6P).

**AG**

AG is an objective sleep variable measurement method with high reliability and validity that is used to assess the circadian rhythm [26]. Participants wore an AG (Actiwatch-2, Philips Respironics, Cambridge, MA, USA) on their non-dominant wrist for about 1 week. Data were stored in 30-second epochs. Total sleep time (TST), sleep latency (SL), sleep efficiency (SE; defined as the ratio between TST and time spent in bed), wake after sleep onset (WASO), and number of awakenings (NOAs) were analyzed using the AG activation data.

**nPSG**

nPSG was performed to evaluate objective sleep parameters and exclude other sleep disorders. We used Neuro system and Profusion EEG software (ver. 3.4; Compumedics, Abbotsford, VIC, Australia) for nPSG recordings in a standard manner and continuously measured electroencephalogram, electrooculogram, electrocardiogram, chin and limbs electromyogram, oral and nasal airflow, snoring sounds, abdominal movement, chest and abdominal movement, body position, and arterial oxygen saturation during night. The TST, WASO, SL, SE, apnea-hypopnea index (AHI), and periodic limb movement index (PLMI) were analyzed. The sleep stages and events during sleep were scored and sleep parameters were calculated based on the American Academy of Sleep Medicine (AASM) criteria [27].

**Statistical analysis**

Data were analyzed using SPSS (ver. 25, IBM Corp., Armonk, NY, USA). The Mann–Whitney U-test was conducted to investigate differences in variables between the OSA and NC groups because the data failed to meet the normality assumption. Spearman’s rho correlation analysis was performed to analyze the relationships among the intermediate and final hexosamine pathway metabolites, sleep parameters, and cognitive function indicators in participants with symptoms of OSA. Mediation analysis was performed with the PROCESS macro [28]. Statistical significance was accepted at p<0.05.

**RESULTS**

**Demographic and baseline clinical characteristics**

We divided participants into a severe OSA group (30≤AHI) and an age-matched NC group (AHI≤15) for the group comparison. Among all participants, six whose age did not match well or whose sleep apnea severity was not clearly classified were excluded from the group comparison. Nineteen participants diagnosed with OSA (5≤AHI) were included in the correlation and mediation analysis. The demographic characteristics of the participants are listed in Table 1.

**Comparison of variables between the OSA and NC groups**

Table 2 shows the differences in objective sleep variables, levels of hexosamine pathway metabolites, and CAT results between the OSA and NC groups. The results, except for AHI, average and minimum O2 saturationm, and PLMI were not significant (p>0.05).

**Correlation analysis of sleep parameters with cognitive performance**

A Spearman’s correlation analysis between sleep variables and CAT results was performed in the OSA patients (Table 3). No significant correlation of nPSG variables was observed (p>0.05). Regarding AG variables, TST was positively correlated with OE (r=0.67, p<0.05) and the mean RT of ISAT (r=0.67, p<0.05). SE was negatively correlated with the CE of ISAT (r=-0.82, p<0.01) and positively correlated with the mean RT of ISAT and DAT (ISAT: r=0.92, p<0.01, DAT: r=0.82, p<0.01). NOA had a positive correlation with CE (r=0.92, p<0.01) and a negative correlation with the mean RT of ISAT (r=-0.80, p<0.01).

**Correlation analysis of sleep parameters with HBP pathway metabolites**

A Spearman’s correlation analysis between sleep variables and hexosamine pathway metabolites was performed in the OSA patients (Table 4). Regarding nPSG variables, TST had a positive cor-
Relation of Hexosamine Pathway with Sleep and Cognition

Spearman’s correlation analysis of hexosamine pathway metabolites and CAT results was performed in the OSA patients (Table 1). The table shows that:

- GlcNAc1P and GlcNAc6P are negatively correlated with F6P and G6P (r=-0.70, p<0.01; GlcNAc1P and GlcNAc6P: r=-0.64, p<0.05). There was a positive correlation with GlcNAc1P and GlcNAc6P and UDP-GlcNAc (GlcNAc1P and GlcNAc6P: r=0.59, p<0.05; UDP-GlcNAc: r=0.65, p<0.01). WASO was negatively correlated with UDP-GlcNAc (r=-0.61, p<0.05). In terms of AG variables, SL was negatively correlated with GlcNAc1P and GlcNAc6P (r=-0.69, p<0.05). SE was positively correlated with the levels of GlcNAc1P and GlcNAc6P (r=0.72, p<0.05). NOA was negatively correlated with GlcNAc1P and GlcNAc6P (r=-0.75, p<0.05).

Correlation analysis of HBP pathway metabolites with cognitive performance

A Spearman’s correlation analysis of hexosamine pathway metabolites and CAT results was performed in the OSA patients (Ta-
levels of hexosamine pathway metabolites mediated the relation-
chip between sleep disturbances, cognitive function, and HBP activ-
ation in middle- and older-aged adults with/without OSA. Sleep
apnea is linked to alterations in, and may suppress, glucose me-
tabolism [29,30]. Regarding the underlying mechanism, Punjabi
and Polotsky [31] conjectured that sleep fragmentation and hy-
poxemia during sleep alter the hypothalamic-pituitary-
adrenal and somatotropic axis, thus affecting glucose tolerance.
However, we observed no significant difference of hexosamine
pathway metabolites between the OSA and NC groups. The rea-
son for this might be the small sample size and/or ethical limita-
tions of human studies. Whereas prior animal studies involved ex-
treme hypoxic conditions [24,32], in this human study, OSA patients
experiencing intermittent hypoxia were recruited. We classified
participants into a severe OSA group and a relatively healthy NC
group, but the experimental conditions might not have been suf-
cient to produce the expected difference.

Participants who slept longer missed more responses (i.e., OE) and
took longer to provide the correct answer. Since there was no signif-
icient correlation with CE, it may just have been that they ex-
ceeded the given time to respond carefully. Those who slept with
high efficiency responded better to target stimuli and took longer
to provide the correct answer, and those who woke less frequent-
ly during sleep responded better to target stimuli. Given that mean
RT and CE revealed opposite correlations, it can be supposed that
individuals who have relatively low sleep quality might tend to re-
spond more impulsively in cognitive judgments. This is support-
ed by previous reports of significant associations between impul-
sivity and shorter TST and lower SE as measured by AG [33], and
shorter sleep duration based on self-reported questionnaires [34].
However, because cognitive change usually occurs over a long pe-
riod and this was not a longitudinal study, the relationship needs
to be confirmed.

Participants who slept less, took more time to fall asleep, had
less efficient sleep, and were awake for more time and more often
during sleep had lower levels of HBP metabolites. Also, those with
lower levels of HBP metabolites made fewer errors in cognitive
judgments. Alzheimer’s disease, which is accompanied by sleep distur-
bance, results in low UDP-GlcNAc levels [21,35]. Lee et al. [24]
reported that O-GlcNAcylation in the zebrafish brain decreased as
sleep deprivation time increased, demonstrating impaired learn-
ing and memory effects. That result implied that sleep dis-
turbance precedes metabolic changes and cognitive decline.

**DISCUSSION**

In the current study, we examined the associations between
objective sleep variables, CAT results, and the levels of hexos-
amine pathway metabolites in blood, to investigate the relation-
ships among sleep disturbances, cognitive function, and HBP ac-
tivation in middle- and older-aged adults with/without OSA. Sleep
apnea is linked to alterations in, and may suppress, glucose me-
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sleep deprivation time increased, demonstrating impaired learn-
ing and memory effects. That result implied that sleep dis-
turbance precedes metabolic changes and cognitive decline.

**Table 2. Differences in variables between the OSA and NC groups (n=18)**

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The GlcNAc1P and GlcNAc6P levels were negatively correlated with CE of ISAT (r=-0.71, p<0.05).

**Mediating effect of HBP activation between sleep parameters and cognitive performance**

A mediation analysis was performed to determine whether the
levels of hexosamine pathway metabolites mediated the relation-
ship between sleep variables and CAT results. We tested two mod-
els using significantly correlated variables for both all participants
and the OSA symptom group. In the first model, SE measured by
AG was used as the independent variable, CE of ISAT as a depen-
dent variable, and GlcNAc1P and GlcNAc6P as mediator vari-
ables. In the second model, NOA measured by AG was used as the
independent variable, CE of ISAT as a dependent variable, and
GlcNAc1P and GlcNAc6P as mediator variables. No significant
mediating effect was observed (p>0.05).
Table 3. Spearman correlation between and sleep and cognitive variables with OSA patients (n=19)

| Variables 1 | 2       | 3       | 4       | 5       | 6       | 7       | 8       | 9       | 10      | 11      | 12      | 13      | 14      | 15      | 16      | 17      | 18      | 19      | 20      |
|-------------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|
| nPSG        |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |
| 1. TST      | 1       |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |
| 2. SL       | -0.15   | 1       |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |
| 3. SE       | 0.56*   | -0.55*  | 1       |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |
| 4. WASO     | -0.46*  | 0.58†   | -0.98†  | 1       |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |
| 5. AHİ      | -0.04   | -0.07   | -0.11   | 0.04    | 1       |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |
| 6. Avg O₂   | 0.10    | 0.17    | 0.07    | 0.00    | -0.87†  | 1       |         |         |         |         |         |         |         |         |         |         |         |         |         |         |
| 7. Min O₂   | -0.01   | 0.22    | -0.07   | 0.10    | -0.76†  | 0.74†   | 1       |         |         |         |         |         |         |         |         |         |         |         |         |         |
| Actigraphy  |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |
| 8. TST      | -0.17   | 0.02    | -0.42   | 0.44    | -0.38   | 0.17    | 0.04    | 1       |         |         |         |         |         |         |         |         |         |         |         |         |
| 9. SL       | -0.40   | -0.01   | -0.20   | 0.18    | -0.36   | 0.15    | -0.10   | 0.37    | 1       |         |         |         |         |         |         |         |         |         |         |         |
| 10. SE      | 0.22    | -0.14   | 0.05    | -0.04   | -0.04   | 0.06    | 0.35    | -0.56*  | 1       |         |         |         |         |         |         |         |         |         |         |         |
| 11. WASO    | -0.18   | 0.40    | -0.13   | 0.17    | 0.04    | -0.09   | -0.06   | 0.05    | 0.49    | -0.68*  | 1       |         |         |         |         |         |         |         |         |         |
| 12. NOA     | -0.09   | -0.04   | -0.02   | -0.03   | 0.05    | 0.07    | 0.00    | -0.36   | 0.36    | -0.84†  | 0.47    | 1       |         |         |         |         |         |         |         |         |
| ISAT        |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |
| 13. CE      | -0.11   | 0.12    | 0.12    | -0.14   | -0.09   | 0.32    | 0.22    | -0.46   | 0.27    | -0.82†  | 0.26    | 0.92†   | 1       |         |         |         |         |         |         |         |
| 14. OE      | 0.14    | 0.20    | -0.16   | 0.23    | -0.27   | 0.21    | 0.16    | 0.67*   | 0.49    | 0.24    | -0.03   | -0.18   | 0.34    | 1       |         |         |         |         |         |         |
| 15. RT      | -0.07   | 0.07    | -0.20   | 0.22    | -0.13   | -0.08   | 0.18    | 0.67*   | -0.20   | 0.92†   | -0.22   | -0.80†  | -0.78†  | -0.08   | 1       |         |         |         |         |         |
| 16. d’      | 0.16    | -0.08   | -0.05   | 0.03    | 0.28    | -0.31   | -0.22   | -0.07   | -0.58   | 0.42    | -0.07   | -0.52   | -0.85†  | -0.71†  | 0.54    | 1       |         |         |         |         |
| DAT         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |
| 17. CE      | -0.13   | -0.29   | 0.45    | -0.41   | -0.17   | 0.24    | -0.15   | -0.31   | 0.40    | -0.08   | 0.03    | 0.07    | 0.19    | -0.10   | -0.18   | 0.16    | 1       |         |         |         |
| 18. OE      | 0.13    | 0.34    | -0.41   | 0.43    | 0.01    | 0.07    | 0.14    | 0.42    | -0.18   | -0.09   | 0.25    | 0.08    | 0.28    | 0.47    | -0.28   | -0.29   | -0.71†  | 1       |         |         |
| 19. RT      | 0.53    | 0.23    | 0.00    | 0.04    | 0.34    | -0.34   | -0.14   | 0.47    | -0.35   | 0.82†   | -0.12   | -0.60   | -0.46   | 0.23    | 0.48    | 0.35    | -0.25   | 0.13    | 1       |
| 20. d’      | -0.06   | -0.36   | 0.41    | -0.43   | 0.00    | -0.11   | -0.10   | 0.15    | 0.40    | -0.37   | -0.38   | -0.47   | -0.48   | 0.47    | 0.42    | 0.53    | -0.94†  | 0.02    | 1       |

*p<0.05, †p<0.01. OSA: obstructive sleep apnea, nPSG: nocturnal polysomnography, ISAT: inhibition-sustained attention test, DAT: divided attention test, TST: total sleep time, SL: sleep latency, SE: sleep efficiency, WASO: wake after sleep onset, AHİ: apnea hypopnea index, Avg O₂: average O₂ saturation, Min O₂: minimum O₂ saturation, NOA: number of awakenings, CE: commission errors, OE: omission errors, RT: mean response time of correct responses, d’: sensitivity coefficient
Based on the prior studies, we expected that levels of metabolites would have a mediating effect between sleep variables and cognitive performance. However, our testing of models in which each variable showed significant correlations indicated that the mediating effects were not significant. This might have been because the number of samples was not sufficient to perform a mediation analysis.

**Limitations**

This study had several limitations. First, the sample size was small. The study should be replicated with a larger number of participants to verify whether the small sample size affected significance. Second, the correlations and cross-sectional design cannot explain causality or directionality between variables. Third, we used only attention tests as a measure of cognitive function and conducted them only once. A further study needs to evaluate longer-term cognitive tests. Fourth, sleep may not have been accurately assessed. One-night nPSG is the most accurate test for measuring sleep parameters by EEG, but does not always represent those of usual sleep. AG is assumed to represent sleep over 7–10 days, although it is less accurate. We used both to obtain more accurate results. Fifth, it is not clear whether blood collected from peripheral veins reflects molecular changes in the brain. Previous studies used the brains of deceased patients or animals to analyze levels of hexosamine pathway metabolites. This methodological difference may have affected the results. Finally, the prevalence of OSA in this study was high since we recruited mid- and older-aged adults, and the criteria of the NC group were defined more generously (AHI≤15) than the clinical criteria (AHI<5). Future studies should be designed in consideration of the points.

**Conclusion**

This is the first study to directly investigate the associations among objective sleep variables, HBP activation, and cognitive function in both OSA patients and healthy controls. The OSA and NC groups showed no significant differences. Sleep parameters, cognitive function, and HBP activation level showed significant relationships. Participants with high sleep quality generally reported more slowly, gave more correct answers in the cognitive test, and had higher levels of hexosamine pathway metabolites. Those who had higher levels of metabolites made fewer errors in the cognitive test.

**Table 4. Spearman correlation between hexosamine pathway metabolites and sleep parameters with OSA patients (n=19)**

<table>
<thead>
<tr>
<th>Variables</th>
<th>1</th>
<th>2</th>
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<th>4</th>
<th>5</th>
<th>6</th>
<th>7</th>
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<th>11</th>
<th>12</th>
<th>13</th>
<th>14</th>
<th>15</th>
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</thead>
<tbody>
<tr>
<td>Hexosamine pathway metabolites</td>
<td>1. F6P, G6P</td>
<td>1</td>
<td>2. GlcNAc1P, GlcNAc6P</td>
<td>0.40</td>
<td>1</td>
<td>3. UDP-GlcNAc</td>
<td>0.80*</td>
<td>0.39</td>
<td>1</td>
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</tr>
<tr>
<td>nPSG</td>
<td>4. TST</td>
<td>0.31</td>
<td>0.33</td>
<td>0.65*</td>
<td>1</td>
<td>5. SL</td>
<td>-0.70*</td>
<td>-0.64*</td>
<td>-0.50</td>
<td>-0.15</td>
<td>1</td>
<td>6. SE</td>
<td>0.51</td>
<td>0.59*</td>
<td>0.65*</td>
</tr>
<tr>
<td>7. WASO</td>
<td>-0.51</td>
<td>-0.48</td>
<td>-0.61*</td>
<td>-0.46*</td>
<td>0.58†</td>
<td>-0.98*</td>
<td>1</td>
<td>8. AHI</td>
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<td>-0.04</td>
<td>-0.07</td>
<td>-0.11</td>
<td>0.04</td>
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<td>Actigraphy</td>
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<td>0.10</td>
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<td>0.02</td>
<td>-0.42</td>
<td>0.44</td>
<td>-0.58</td>
<td>0.17</td>
<td>0.04</td>
<td>1</td>
<td>12. SL</td>
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<td>0.69*</td>
</tr>
<tr>
<td>13. SE</td>
<td>0.03</td>
<td>0.72*</td>
<td>0.05</td>
<td>0.22</td>
<td>-0.14</td>
<td>0.05</td>
<td>-0.04</td>
<td>-0.01</td>
<td>-0.04</td>
<td>0.06</td>
<td>0.35</td>
<td>-0.56*</td>
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<tr>
<td>14. WASO</td>
<td>-0.45</td>
<td>-0.59</td>
<td>-0.32</td>
<td>-0.18</td>
<td>0.40</td>
<td>0.13</td>
<td>-0.17</td>
<td>0.04</td>
<td>-0.09</td>
<td>-0.06</td>
<td>0.05</td>
<td>0.49</td>
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<td>15. NOA</td>
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<td>-0.03</td>
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<td>0.07</td>
<td>0.00</td>
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<td>0.36</td>
<td>-0.84*</td>
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Table 5. Spearman correlation between hexosamine pathway metabolites and cognitive variables with OSA patients (n=19)

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<tr>
<td>1. F6P, G6P</td>
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<tr>
<td>2. GlcNAc1P, GlcNAc6P</td>
<td>0.40</td>
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<tr>
<td>3. UDP-GlcNAc</td>
<td>0.80*</td>
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<tr>
<td>Inhibition-sustained attention test</td>
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<tr>
<td>4. CE</td>
<td>0.07</td>
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<td>5. OE</td>
<td>-0.07</td>
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<tr>
<td>6. RT</td>
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<td>0.47</td>
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<td>-0.78*</td>
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<td>7. d’</td>
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<td>0.61</td>
<td>-0.01</td>
<td>-0.85*</td>
<td>-0.71*</td>
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<td>Divided attention test</td>
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<td>8. CE</td>
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<td>9. OE</td>
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<td>0.47</td>
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<td>-0.29</td>
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<td>10. RT</td>
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<td>11. d’</td>
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<td>0.40</td>
<td>-0.12</td>
<td>-0.47</td>
<td>0.48</td>
<td>0.47</td>
<td>0.42</td>
<td>0.53</td>
<td>-0.94*</td>
<td>0.02</td>
<td>1</td>
</tr>
</tbody>
</table>

*p<0.05, †p<0.01. OSA: obstructive sleep apnea, F6P: fructose 6-phosphate, G6P: glucose 6-phosphate, GlcNAc1P: N-acetyl-D-glucosamine 1-phosphate, GlcNAc6P: N-acetylglucosamine-6-phosphate, UDP-GlcNAc: uridine diphosphate N-acetylglucosamine, CE: commission errors, OE: omission errors, RT: mean response time of correct responses, d’: sensitivity coefficient

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Conflicts of Interest
The authors have no potential conflicts of interest to disclose.

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